SUPPLEMENTAL MATERIAL

The pro-angiogenesis effect of miR33a-5p/Ets-1/DKK1 signaling in ox-LDL induced HUVECs

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Figure S1. DKK1 did not participate in the proliferation of ox-LDL-treated HUVECs, and DKK1induced angiogenesis did not occur via the canonical Wnt/ β -catenin pathway. (A) HUVECs were transiently transfected with negative control (NC) and DKK1 siRNA (si-DKK1) for 24h and then treated with 150 µg/ml ox-LDL for 6h. The percentage of Edu (+) cells was calculated and quantified. Bars indicate 50 µm. (B) Cells were pretreated with DMSO or IM-12 (3 µM) for 1 h and then transfected with lenti-DKK1. Western blot to quantify the VEGF-A, VEGFR-2, MMP2, MMP9 and β -catenin protein levels. The data are shown as the mean±SEM. n=6. **P*<0.05 vs.NC.



-2029 -2023 Binding Site

Figure S2. c-jun promotes the expression of DKK1 by binding with a key positive regulatory region of the human DKK1 promoter. (A-B) HUVECs treated with ox-LDL (150 μ g/ml) for various lengths of time (0h, 1h, 3h, and 6h). Western blot and qRT-PCR to quantify the Ets-1, c-jun or c-fos protein and mRNA levels. (C) The immunofluorescence localization of Ets-1, c-jun or c-fos in HUVECs treated with ox-LDL (150 μ g/ml) for 6h. Bars indicate 20 μ m. (D-E) HUVECs were transiently transfected with NC or c-jun siRNA (si-c-jun) for 24h and then treated with 150 μ g/ml ox-LDL for 6h. Western Blot and qRT-PCR were used to quantify the DKK1 protein and mRNA levels. (F-G) The luciferase activities were analyzed: (F) Full-length DKK1 promoter in 293T cells transfected with c-jun siRNA. (G) Rough characterization of the DKK1 promoter using a serial fragment from -2034 to -234bp. pcDNA3.0+P0, pcDNA3.0-c-jun +P0-P9 were transfected into 293T cells. (H) The prediction of c-jun binding sites in this region (-2034 ~ -1834bp) of the DKK1 promoter: Binding site (-2029 to -2023bp). The data are shown as the mean±SEM. n=6. **P*<0.05 vs. the untreated group or NC; #*P*<0.05 vs. NC+ox-LDL or P0+ PCDNA3.1-c-jun.



Figure S3. MiRNA participated in the upregulation of Ets-1 and DKK1 in ox-LDL-induced HUVECs, but miR33a-5p did not directly targeted DKK1. (A) HUVECs were transiently transfected with NC or Dicer siRNA for 24h and then treated with 150 μ g/ml ox-LDL for 6h. Western blotting was used to quantify the Ets-1 and DKK1 protein levels. (B) Possible binding sites for miR33a-5p in the DKK1 3'-UTR, as predicted. A miR target reporter luciferase assay was performed after the miR33a-5p mimic and inhibitor were delivered to 293T cells. The results were normalized to data obtained from an assay with Renilla luciferase. The data are presented as the mean ± SEM. n=6. **P*<0.05 vs. the NC group or the control group; #*P*<0.05 vs. NC+ ox-LDL or P0+ PCDNA3.1-c-jun.



Figure S4. miR33a-5p/Ets-1 participated in the angiogenic effects in HUVECs via assistance of CBP/P300.

(A-C) HUVECs were cotransfected with an miR33a inhibitor and Ets-1 siRNA. (D-F) HUVECs were pretreated with DMSO or a CBP/P300 inhibitor (25 μ M) for 1h after transfection with lenti-Ets-1(Ets-1). Representative images and quantification of cell migration in Transwell assays. The cell counts on the bottom of the Transwell are shown here: (A, D) Representative images and quantification of cell migration in the transwell are shown here. Bars indicate 200 μ m. (B, E) Representative images and quantification of cell migration in the in vitro scratch wound assay (relative to 0 h) were obtained at 0h and 24h post-wounding. Bars indicate 100 μ m. (C, F) Representative image and quantification of the tube length (% of control). Bars indicate 100 μ m. The data are presented as the mean±SEM. n=6. **P*<0.05 vs. the NCI group or the DMSO group; #*P*<0.05 vs. the miR33a-5p inhibitor or the DMSO+Ets-1 group.

Major Resources Table

Animals (in vivo studies)

Species	Vendor or Source	Background Strain	Sex
ApoE-/- mice	Beijing HFK Bioscience Co.,Ltd.	C57BL/6J	male

Antibodies

Target	Vendor or	Catalog #	Working concentration
antigen	Source		
β-actin	Cell Signaling	3700	WB(diluted1:1000)
	Technology		
DKK1	Abcam	ab109416	WB(diluted 1:500); IHC(diluted1:100)
P300	Abcam	ab14984	WB(diluted 1:500);
CBP	Abcam	ab2832	WB(diluted 1:500); IP(diluted1:50)
c-fos	Abcam	ab214672	WB(diluted 1:1000); IF(diluted1:200)
c-jun	Abcam	ab32137	WB(diluted 1:2000); IF(diluted1:200)
Ets-1	Abcam	ab96478(mice antibody)	WB(diluted 1:1000); IF(diluted1:400);
			ChIP (diluted 1:50)
Ets-1	Cell Signaling	14069(rabbit antibody)	WB(diluted1:1000);IP (diluted 1:50)
	Technology		
IgG1	Cell Signaling	3900	IP (diluted 1:50)
	Technology		
CD31	Abcam	ab281583	IHC(diluted1:50)
VEGF-A	Abcam	ab52917	WB(diluted 1:10000); IHC(diluted1:250)
VEGFR-2	Abcam	ab39638	WB(diluted 1:1000); IHC(diluted1:100)
MMP2	Abcam	ab37150	WB(diluted 1:1000); IHC(diluted1:50)
MMP9	Abcam	ab38898	WB(diluted 1:1000); IHC(diluted1:50)
PI3K	Cell Signaling	4249	WB(diluted1:1000)
	Technology		
CKAP4	Abcam	ab152154	WB(diluted1:10000)
β-catenin	Cell Signaling	8480	WB(diluted1:1000)
	Technology		

DNA/cDNA Clones

Clone Name	Sequence
β-actin primer	Forward: CGTGCGTGACATTAAGGAGA
	Reverse: CACCTTCACCGTTCCAGTTT
DKK1 primer	Forward: ATAGCACCTTGGATGGGTATTCC
	Reverse: CTGATGACCGGAGACAAACAG
Ets-1 primer	Forward:TACACAGGCAGTGGACCAATC
	Reverse: CCCCGCTGTCTTGTGGATG
c-fos primer	Forward: CACTCCAAGCGGAGACAGAC
	Reverse: AGGTCATCAGGGATCTTGCAG
c-jun primer	Forward: TCCAAGTGCCGAAAAAGGAAG
	Reverse: CGAGTTCTGAGCTTTCAAGGT
U6 primer	Forward: CAGCACATATACTAAAATTGGAAGG
	Reverse: ACGAATTTGCGTGTCATCC

GGA
GGA
A
GGA
CA
GGA
CA
CGA
CA

Cultured Cells and reagents

Name	Source and working concentration
HUVECs	ScienCell
НЕК-293Т	American Type Culture Collection
ox-LDL	Yiyuan Biotechnologies(Guangzhou, China), 150 µg/ml
CBP/P300 inhibitor	Abcam, ab142163,2ug/ml
IM-12	Selleck, S7566, 3 μM
740 Y-P	Selleck, S7865, 50 µg/ml