## **Supplementary Figures:**



Figure S1. EC-specific Cas9-expressing mouse exhibited a normal brain vasculature.

(A) The strategy used for generating  $Tie2^{Cas9}$  mice. (B) Sulfo-NHS-LC-Biotin tracer extravasation assay revealed no significant differences in BBB integrity in brain sections of control and  $Tie2^{Cas9}$  mice. Scale bar, 40 µm. (C-E) Quantification of the cerebral capillary length (n = 3 mice), microvessel diameters (n = 5 mice), and branch points (n = 4 mice), showing a comparable brain vasculature in control and  $Tie2^{Cas9}$  mice. Data are means ± SEM. (NS = not significant, two-tailed unpaired *t*-test).



Figure S2. AAV-BR1 specifically transduces ECs in brain.

(A-F) Immunostaining of tdTomato (red), CD31 (green) and DAPI (blue) from brain (A), liver (B), spleen (C), kidney (D), stomach (E) and heart (F) 4 weeks after vector injection, showing that AAV-BR1-tdTomato specifically transduces the brain ECs. Scale bar, 50 μm.



Figure S3. In vivo genome editing of brain ECs using AAV-BR1-CRISPR system.

(A) T7E1 analysis on target sites of PCR-amplified genomic DNA in lungs, spinal cord, liver, and kidney from control and  $Tie2^{Cas9}$  mice 4 weeks after AAV-BR1-sg*Ctnnb1*-tdTomato injection. Lane I was loaded with a molecular weight marker (100 bp ladder). (B) T7E1 analysis of the top 5 potential OT sites of PCR-amplified genomic DNA of brain ECs isolated from  $Tie2^{Cas9}$  mice 4 weeks after AAV-BR1-sg*Ctnnb1*-tdTomato injection. Lane I was loaded with a molecular weight marker (100 bp ladder). (C) Sulfo-NHS-LC-Biotin tracer (green) injection generated BBB defects, mainly in tdTomato positive area. Scale bar, 30 µm. (D) Immunostaining of CLAUDIN-5 (red), PLVAP (green) and CD31 (purple) showing downregulation of CLAUDIN-5, and upregulation of PLVAP in  $Tie2^{Cas9}$  mice at P60 (4 weeks after AAV-BR1-sg*Ctnnb1*-tdTomato treatment). Scale bar, 50 µm.

## **Supplementary Table:**

SgRNA oligos and primers for genotyping, T7E1 analysis, DNA sequencing, and qRT-PCR analysis.

Name	Sequence(5'-3')	Purpose
Tie2 Cre fwd	GAAGGGCAAGATGGATAGGGC	mouse genotyping
Tie2 Cre rev	GCATCGACCGGTAATGCAGGC	mouse genotyping
Cas9 Tg-1	GAGGCAGGAAGCACTTGCTCT	mouse genotyping
Cas9 Tg-2	TGGCGTTACTATGGGAACATACG	mouse genotyping
Cas9 Tg-3	TACACCTGTTCAATTCCCCTGC	mouse genotyping
sgCON fwd	CACCGAGCTCGCCATGTCGGTTCTC	In vitro sgRNA
sgCON rev	CGTGAACCGACATGGCGAGCTCAAA	In vitro sgRNA
Ctnnb1 sgRNA1 fwd	CACCGGCGTGGACAATGGCTACTCA	In vitro and in vivo sgRNA
Ctnnb1 sgRNA1 rev	CTGAGTAGCCATTGTCCACGCCAAA	In vitro and in vivo sgRNA
Ctnnb1 sgRNA2 fwd	CACCGTGACCTGATGGAGTTGGACA	In vitro sgRNA

Ctnnb1 sgRNA2 rev	CTGTCCAACTCCATCAGGTCACAAA	In vitro sgRNA
Ctnnb1 sgRNA3 fwd	CACCGGTTGGACATGGCCATGGAGC	In vitro sgRNA
Ctnnb1 sgRNA3 rev	CGCTCCATGGCCATGTCCAACCAAA	In vitro sgRNA
sgRNA1 fwd	AGCACCGTATGCCTACAATCT	T7E1 primer
sgRNA1 rev	CATCTTCTTCCTCAGGGTTGC	T7E1 primer
sgRNA 2 fwd	CTCTTCCCTTCTGCACACTAC	T7E1 primer
sgRNA 2 rev	GGACATTAGTGGGATGAGCA	T7E1 primer
sgRNA 3 fwd	CTCTTCCCTTCTGCACACTAC	T7E1 primer
sgRNA 3 rev	GGACATTAGTGGGATGAGCA	T7E1 primer
Sobp fwd	GTGAGACGACTAGACACGCTTAG	Ctnnb1 OT1 T7E1
Sobp rev	GCCTGTCTATCTTATTGCTTTAGC	Ctnnb1 OT1 T7E1
<i>Llgl2</i> fwd	CCTACATTGACTTCGGAGAGG	Ctnnb1 OT2 T7E1
Llgl2 rev	GTCAGGGACAATGTGATTAAGAAC	Ctnnb1 OT2 T7E1
<i>Gm10851</i> fwd	CACATACACACATGAGCACATG	Ctnnb1 OT3 T7E1
<i>Gm10851</i> rev	CTCAGTGAGGGACTGTCTGAC	Ctnnb1 OT3 T7E1
<i>Gtpbp3</i> fwd	CATGAGTCACAGCTGTATTCACTT	Ctnnb1 OT4 T7E1
<i>Gtpbp3</i> rev	CTCTAGATCAACTGAGCAAAGCT	Ctnnb1 OT4 T7E1
<i>Gm26812</i> fwd	GAGTATTGACCTAGCACCTGATAG	Ctnnb1 OT5 T7E1
<i>Gm26812</i> rev	CACGTGGTGAGAGCTGAATC	Ctnnb1 OT5 T7E1
	AGATCGGAAGAGCACACGTCTGAACTCCA	
Ctnnb1 fwd	GTCACNNNN	DNA sequencing
	GTATAGGTAGCAGAATCACGG	

Ctnnb1 rev	TGCTGATTATTTCACCAAGCCNNNNAGATC	
	GGAAGAGCGTCGTGTAGGGAAAGAGTG	DNA sequencing
Ctnnb1 exon 9 fwd	GGTGCTATTCCACGACT	qRT-PCR
Ctnnb1 exon 10 rev	CCCTTCTACTATCTCCTCC	qRT-PCR
GAPDH fwd	TGCCCAGAACATCATCCCT	qRT-PCR
GAPDH rev	GGTCCTCAGTGTAGCCCAAG	qRT-PCR

\*NNNN = index from Illumina TruSeq Small RNA Sample Prep Kit