# Supplementary Material

# Chemical-proteomics Identify Peroxiredoxin-1 As An Actionable Target In Triple-negative Breast Cancer

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# Supplementary Material:

- Supplementary Methods
- Supplementary Tables
- Supplementary Figures

#### **Molecular Docking**

The crystal structure of the dimeric Prdx1 in complex with sulfiredoxin (PDB ID: 3HY2) [1] was downloaded from the RCSB database [2]. We choose to use this complex structure as the basis for our modeling studies because of its reasonable atomic resolution. We used Maestro's Protein Preparation Wizard [3, 4] to hone the structure. Prime [5-7] was used to add missing atoms and residues to the protein structure and complete the incomplete parts of the loops. After the original hydrogen atoms were deleted, the bond orders of the amino acid residues and ligands were readjusted. The degree of protonation was altered to achieve the desired pH value. Only water molecules within 5 Å from ligands and active sites were maintained to save calculation time. Hydrogen bonds were sampled at pH 7.4 using PROPKA [8], with the water molecule orientations in the active site altered to achieve this value. The OPLS3e force field [9] was used to geometrically refine the protein complex until an RMSD of 0.3 Å was reached via restricted reduction with heavy atom convergence, therefore resolving any remaining steric conflicts. The receptor grids were prepared using the OPLS3e force field. SiteMap [10-12] is utilized for locating druggable Prx1 areas. The sensitivity of the receptor was enhanced by boosting the Van der Waals radii of its non-polar atoms by a factor of 1. A requirement of 0.25 or less for the absolute partial atomic charge for a non-polar atom was considered. The centroids of the ligand binding regions have been used to establish the centers of 20 Å<sup>3</sup> grids. No constraints were imposed on rotations in any of the binding locations, and no receptor grids were used. Glide [13 - 16] was used to dock the fully prepared ligands into the receptor grids in Maestro with XP docking precision. Setting the scaling factor for Van der Waals radii of the ligand's nonpolar atoms to 0.8 and the absolute partial atomic charge cut-off to 0.15 allowed for the exploration of a softer potential during docking simulations. All poses were minimized following docking.

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## Table S1. Antioxidants information.

Product	Company	Cat#	Concentration	
Catalase from bovine liver	Sigma-Aldrich	C1345-10G	1000 U/ml	
N-acetyl-L-cysteine (NAC)	Sigma-Aldrich	A7250-25G	2 mM	
L-glutathione reduced (GSH)	Sigma-Aldrich	G6013-5G	1 mM	
<b>α</b> -tocopherol (vitamin E)	Sigma-Aldrich	258024-5G	20 µM	
Superoxide Dismutase (SOD)	Sigma-Aldrich	S5395-15KU	200 U/ml	

### Table S2. Antibodies information.

Cell Signaling Technology							
Antibody	Cat#	Lot#	Lot# Dilution				
p-Akt (Ser473)	9271	14	1:1,000	AB_329825			
Akt	9272	22	1:1,000	AB_329827			
Bax	2772	11	1:1,000	AB_10695870			
Bcl-xL	2764	11	1:1,000	AB_2228008			
Bcl-2	4223	6	1:1,000	AB_1903909			
BID	2002	5	1:1,000	AB_10692485			
p-ERK (T202/Y204)	9101	26	1:1,000	AB_331646			
ERK1/2	9102	27	1:1,000	AB_330744			
p-SAPK/JNK (T183/Y185)	9251	21	1:1,000	AB_331659			
Prdx1	8499	1	1:1,000	AB_10950824			
p-p38 MAPK (T180/Y182)	9216	27	1:2,000	AB_331296			
р38 МАРК	9212	23	1:1,000	AB_330713			
p21 Waf1/Cip1	2947	10	1:1,000	AB_823586			
p-Stat3 (8727)	9134	20	1:1,000	AB_331589			
p-Stat3 (Y705)	9131	30	1:1,000	AB_331586			
Stat3	9139	8	1:2,000	AB_331757			

Wee1	13084	1	1:4,000	AB_2713924				
p-Cdc2 (Y15)	9111	11	1:8,000	AB_331460				
p27	3686S	4	1:4,000 AB_2077850					
	Santa Cruz Bi	otechnology						
Antibody	Cat#	Dilution	RRID					
Cyclin B1	sc-245	F015	1:5,000 AB_627338					
GADPH	sc-166574	I1013	1:10,000	,000 AB_2107296				
JNK 1/3	sc-474	A0109	1:1,000 AB_632383					
Mcl-1	sc-12756	D2617	1:1,000 AB_627915					
β-actin	sc-81178	J1116	1:2,000	AB_2223230				
anti-rabbit IgG-HRP	sc-2357	A0318	1:5,000 AB_628497					
m-IgGк BP-HRP	sc-516102	1:5,000	AB_2687626					
Enzo Life Sciences								
Antibody	Cat#	Dilution	RRID					
Calnexin	ADI-SPA-860-F	12301304	1:40,000	AB_11178981				
Caspase-3	ADI-AAP-113	11021102	1:1,000	AB_10615972				
Caspase-8	ADI-AAM-118-E	4041308	1:1,000	AB_2038943				
Caspase-9	ADI-AAM-139-E	11221306	1:1,000	AB_2038946				
	BioLeg	gend						
Antibody	Cat#	Lot#	Dilution	RRID				
p-H2AX (8139)	613401	B182042	1:5,000	AB_315794				
BD Pharmingen								
Antibody	Cat#	Lot#	Dilution	RRID				
PARP	551024	38180	1:1,000	AB_394008				
Abcam								
Antibody	Cat#	RRID						
Prdx-SO3	ab16830	GR3294252-1	1:2,000	AB_443491				
BD Biosciences								
Antibody	Cat#	Lot#	Dilution	RRID				
Cyclin A	611268	5023928	1:6,000	AB_398796				

EMD Millipore								
Antibody	Cat#	Lot#	RRID					
p-Histone H3 (S10)	06-570	3076467	1:5,000	AB_310177				
Bio-Rad								
Antibody	Cat#	Lot#	Dilution	RRID				
anti-rabbit IgG-HRP	170-6515	L1706515RevF	1:20,000	AB_11125142				
GE Healthcare Life Sciences								
Antibody	Cat#	Lot#	Dilution	RRID				
anti-mouse IgG-HRP	NA931	17179804	1:10,000	AB_772210				



**Figure S1. Effects of taselisib and napabucasin in TNBC cells. A-B**, Effect of taselisib and napabucasin on cell viability, assessed by MTT metabolism at 48 h in MDA-MB-231 (A) and BT-549 cells (B) (n=2). C, Representative images of colony formation after 9 days of treatment with napabucasin and taselisib in MDA-MB-231 cells (n=2).



Figure S2. Effects of transient exposure to CM728 in MDA-MB-231 cells. A, Effect of pulse exposure of 1  $\mu$ M CM728 on cell number (left) and the percentage of trypan blue-negative cells (right) at 48 h (n=2). Cell counting was performed using trypan blue staining. Statistical analyses were performed using one-way ANOVA and Dunnett's multiple comparison test. \*\*\*, P < 0.0001 *versus* control; ###, P < 0.0001 *versus* CM728 sustained-treated cells. **B**, Visualization of cells after a transient exposure to CM728 (1  $\mu$ M). Images were obtained with an inverted phase-contrast microscope at 20x magnification.



**Figure S3. Evaluation of CM728 oral toxicity. A-B,** Body weight of mice orally administered with vehicle (NMP/PEG400/0.1% methylcellulose in water for injection: 5/25/70) or CM728 (10 mg/kg) for 10 days.



**Figure S4. CM728 binds to Prdx1, inducing its dimerization and hyperoxidation. A,** Identification of CM728-binding proteins in MDA-MB-231 lysates previously incubated with an excess of free CM728. Chemical proteomic approaches were carried out as specified in the Materials and Methods section. **B,** Immunoblot of the monomer (M) and dimer (D) forms of Prdx1, or the levels of hyperoxidized peroxiredoxin (SO<sub>2/3</sub>-Prdx) in response to CM728 (1  $\mu$ M) in MDA-MB-231 cells.  $\beta$ -actin was used as a loading control. Densitometry quantification from immunosignal values is shown below the bands. Values are relativized to the loading control. Representative images of two independent experiments are shown.

	MDA-MB-231				BT-549					
	Time with CM728, h				Tir	Time with CM728, h				
	0	3	6	12	24	0	3	6	12	24
p-JNK	_	-	_	-	=		-	-		=
JNK	-	-	-	-	-	_	_	_	_	_
	1.0	1.6	2.0	4.1	10.7	1.0	1.4	1.4	2.2	7.0
р-р38	-	-	-	-	-		-	-	-	-
p38	-	-	-	-	-	=	=	=	=	=
	1.0	3.6	6.7	9.2	19.7	1.0	1.8	2.5	3.8	6.2
p-ERK1/2				-	-	-	-	-	-	-
ERK1/2	-	-	-	-	-	_	_	_	_	-
	1.0	0.4	0.5	1.4	4.2	1.0	2.8	1.4	1.5	2.8
pS <sup>727</sup> -STAT3	1 mar	-	-	-	-	-	-		-	-
pY <sup>705</sup> -STAT3	-	-	and the second	NUMBER	-	-		-	-	
STAT3	_	-	_	_	_	_	-	-	-	-
pS <sup>727</sup> -STAT3/STAT3	1.0	1.3	1.7	1.3	1.3	1.0	1.0	1.1	1.3	1.9
pY <sup>705</sup> -STAT3/STAT3	1.0	1.0	0.3	0.3	0.1	1.0	0.2	0.2	0.4	0.1
pS <sup>473</sup> -Akt	James .	-	-	-	-	-	-		-	
Akt	-	-	-	-	-	-	-	-	-	-
	1.0	2.0	1.6	1.1	0.7	1.0	0.8	0.4	0.3	1.2
β-actin	-	-	-	-		-	-	-	-	-

Figure S5. Phosphorylation of MAPKs, STAT3, and Akt in response to CM728 in TNBC cells. Effect of CM728 (1  $\mu$ M) on JNK, p38 MAPK, ERK1/2, STAT3, and Akt phosphorylation in MDA-MB-231 (left) and BT-549 cells (right).  $\beta$ -actin was used as a loading control. Densitometric quantification from immunosignal values are shown below the bands. Phosphorylated forms of proteins are relativized to the total amount of the corresponding proteins. Values are relativized to the loading control. Representative images of two independent experiments are shown.



Figure S6. CM728-induced cell cycle arrest, DNA damage, and death was blocked by NAC. A-B, Cell cycle was analyzed by propidium iodide staining and flow cytometry in MDA-MB-231 cells that were preincubated with 2 mM NAC for 2 h and then treated with 1  $\mu$ M CM728 for 48 h. A, Representative histogram (left), and percentage of cells in each cycle phase (right); and, B, percentage of subG0 population are shown (n=2) C, Analysis of p21, pS<sup>139</sup>-H2AX and PARP in MDA-MB-231 cells that were preincubated with 2 mM NAC for 2 h and then treated with 1  $\mu$ M CM728 for 24 h.  $\beta$ -actin was used as loading control. Densitometry quantification from immunosignal values relativized to the loading control is shown below the bands. Representative images of two independent experiments are shown. D, Effect of NAC on the CM728-induced apoptosis in MDA-MB-231 cells. The effect of 2 mM NAC (2 h of preincubation) in the absence and presence of 1  $\mu$ M CM728 (48 h of incubation) was studied by flow cytometry after Annexin V/PI double staining (n=2). Representative images of two independent experiments are shown. PI: propidium iodide; An: annexin V. Statistical analyses were performed using two-way ANOVA followed by Tukey's post-hoc test (A) or using one-way ANOVA, and Dunnett's multiple comparison test (B). \*\*\*, P < 0.0001 versus control; ###, P < 0.0001 versus CM728-treated cells.



**Figure S7. CM728 antitumor activity was enhanced by S63845. A,** The cytotoxicity of S63845 was measured by MTT assays after 48 h of incubation in MDA-MB-231 cells (n=2). Statistical analyses were performed with one-way ANOVA, and Dunnett's multiple comparisons test. \*, P < 0.05 versus cells treated with vehicle. **B,** The effect of the indicated concentrations of CM728 alone or in combination with S63845 (0.5  $\mu$ M) was assessed by MTT metabolism after 72 h of treatment in MDA-MB-231 cells (n=2). Statistical analyses were performed with one-way ANOVA, and Dunnett's multiple comparisons test. \*, P < 0.05 versus cells treated with comparisons test. \*, P < 0.05 versus cells treated with CM728 alone; \*\*, P < 0.001 versus cells treated with CM728 alone; \*\*, P < 0.001 versus cells treated with CM728 alone.